

**Special Issue: Second International Conference for Pharmaceutical Sciences (SICPS 2025),
University of Misan, Maysan, 62001, Iraq**

[Journal of Advances in Medical and Biomedical Research | ISSN:2676-6264](#)

The Association of Interferon Lambda 4 (IFNL4) rs12979860 Polymorphism with COVID-19 Outcome

Sara Fakhri Ahmed^{1*}, Muhammed A. H. Aldabagh², Aryan F. Khorshid³,
Dahat Jamal Hawezy³

1. Department, Department of Nursing, Medical Technical Institute, Northern Technical University, Kirkuk, Iraq
2. Department of Microbiology, Collage of Medicine, Al-Nahrain University, Baghdad, Iraq
3. Cardiology Hospital, Sulaimaniyah D.O.H, Sulaimaniyah, Iraq



Article Info

 [10.30699/jambr.33.162.81](https://doi.org/10.30699/jambr.33.162.81)

Received: 2025/10/11;

Accepted: 2025/11/25;

Published Online: 29 Dec 2025;

Use your device to scan and read the article online



*Corresponding author:

Sara Fakhri Ahmed,
Department, Department of Nursing,
Medical Technical Institute, Northern
Technical University, Kirkuk, Iraq

Email: Sara_Fakhria@ntu.edu.iq



Copyright © 2025, This is an original open-access article distributed under the terms of the [Creative Commons Attribution-noncommercial 4.0](#) International License which permits copy and redistribution of the material just in noncommercial usages with proper citation.

ABSTRACT

Background & Objective: Coronavirus Disease 2019 (COVID-19) has a different fatality rate and clinical effects and exhibits a wide range of clinical symptoms, from asymptomatic to critically ill infections that require intensive care units. IFNL4 has shown antiviral activity against RNA viruses. IFNL4 is classified as a type-III interferon the newest addition to the interferon lambda protein family. The receptor of IFNL4 (IFNLR1) is primarily restricted to tissues of epithelial origin. Therefore, interferon lambda proteins may have evolved specifically to protect the epithelium. Genetic polymorphism is one of the factors impacting COVID-19 outcomes. Variants of the IFNL4 gene have been linked to the clinical response to some viral infections. Many studies have found a correlation between the T Allele of IFNL4 (rs12979860) SNP and COVID-19 outcomes. Our goal was to investigate the correlation between such a polymorphism and the severity of COVID-19 infection.

Materials & Methods: A total of 150 positive COVID-19 patients (75 severe, 75 mild/moderate) were enrolled in the present study. The polymorphism was genotyped using TaqMan real-time PCR (polymerase chain reaction).

Results: The present study found the TT genotype of the rs12979860 variants was significantly prevalent in severe patients compared to mild and moderate infection patients, suggesting that this genotype could be a risk factor for COVID-19.

Conclusion: Between the two study groups, there is a statistically significant association between COVID-19 outcome and the T Allele of the IFNL4 (rs12979860) SNP.

Keywords: IFNL4, rs12979860, COVID-19, SARS-CoV-2

1. Introduction

Coronavirus Disease 2019 (COVID-19), which is caused by the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), was found to have a significant fatality rate (1-3). The symptoms and progression of SARS-CoV-2 infection vary greatly among patients, and this variation cannot be attributed to age or pre-existing conditions alone (4). Research increasingly shows that major shifts in cytokine levels in the blood are a key driver of the disease process in severe COVID-19 (5). The cytokine storm is a key factor in triggering the pathological chain of events—including plasma leakage, vascular permeability, and coagulopathy—that leads to life-threatening respiratory

symptoms in COVID-19 (6). The infection progresses when SARS-CoV-2 moves into the lung's air sacs (alveoli) and attaches to type 2 pneumocytes, initiating viral replication. Following replication, the self-destruction of these lung cells causes them to emit inflammatory mediators, which in turn recruit immune cells like macrophages and neutrophils (7).

The stimulated macrophages further secrete cytokines such as IL-1, IL-6 and Tumor Necrotic Factor α (TNF- α). The released cytokines trigger a “cytokine storm”, which stimulates the release of vascular endothelial growth factor (VEGF), monocyte chemoattractant protein-1 (MCP-1), IL-8, and additional IL-6, as well as reduced E-

cadherin expression on endothelial cells causing vasodilation and increased capillary permeability (8). The resulting plasma leakage causes interstitial and alveolar edema. The alveolar fluid decreases surfactant levels, leading to increased surface tension and subsequent alveolar collapse. Radiologically, this edema and collapse manifest as multiple peripheral ground-glass opacities in both lungs, a frequent observation in patients (9). The wide variation in COVID-19 symptom severity suggests that an individual's genetic makeup significantly influences their susceptibility to the virus and the seriousness of their illness (10, 11). Investigations into the role of host genetics in COVID-19 progression have pinpointed numerous susceptibility genes, albeit with differing levels of supporting evidence (12-15). Interferon lambda 4 (IFNL4) gene encodes the IFNL4 protein, which contributes to the innate immune response against viral infection. This protein is categorized as type-III interferons because its signal is through IFNLR1 and IL10R2 (16, 17). Contrary to the widespread expression of IFN- α receptors, IFNLR1 is primarily present in epithelial tissues (18, 19). A human IFNL4 single nucleotide polymorphism (SNP) rs12979860 was associated with good response to treatment of chronic hepatitis C (20, 21) and self-limited HCV infection (22, 23). Also, IFNL4 (rs12979860) has been linked to how SARS-CoV-2 infection progresses, with the T allele that have an influence on clinical progression to severity (24, 25). Our objective was to determine whether the IFNL4 (rs12979860) was associated with the severity of COVID-19 infection.

2. Materials and Methods

Blood samples were collected from 150 COVID-19 patients (75 severe, 75 mild/moderate) who were hospitalized at the internal Teaching Hospital in Sulimaniyah city/ Iraq from October 2022 to March 2023 due to respiratory symptoms and were positive for SARS-CoV-2 (PCR-test). All subjects, were non-vaccinated previously, and there were 83 males (mean age 40 years) and 67 females (mean age 37 years). Each subject signed an informed consent for the genetic study approved by the Ethical Research Committee of the College of Medicine /AL-Nahrain University. DNA was extracted from the blood samples of each subject by using conventional kits addbio genomic DNA extraction kit according to the manufacturer's instructions (addbio, South Korea). IFNL4 (rs12979860) C/T SNP was determined in all the patients by real time PCR genotyping with a TaqMan probe (Fisher Scientific, assay id. C_7820464_10) with Probe sequence of IFNL4 gene rs12979860:

TGAACCAGGGAGCTCCCCGAAGGCG[C/T]
GAACCAGGGTTGAATTGCACTCCGC. The PCR amplification reaction was done according to the manufacturer's instructions using the Add Star Taq master mix PCR kit (addbio, Korea). The PCR reaction

was done by mixing 5 μ L of Green Master Mix and 0.5 μ L of each primer at 10 pmol/L concentration, 0.5 μ L of each probe, 0.5 μ L of MgCl2 and 2 μ L of DNA template. Using nuclease-free water, the capacity was increased to 10 μ L. The thermocycler (Bio-Rad, USA) was set up for an initial denaturation phase of 10 min at 95 °C, followed by 40 cycles of denaturation at 94 °C for 30 s, annealing at 50 °C for 30 s, extension at 72 °C for 40 s, and a final extension phase of 7 min at 72 °C. Detection genotyping kits based on the use of 2 fluorophores. These are based on two probes stained with two different fluorophore dyes, VIC and FAM. The probe stained with FAM indicates the presence of the Mutant Allele (C and T) of IFITM3 rs12252 IFNL4 rs12979860 respectively. The amplification signal for each dye will be considered whenever the detector curve crosses its threshold value. Therefore, samples will be considered to have the wild allele whenever they display a VIC positive signal. By contrast, samples will be considered having a Mutant allele whenever they display a FAM positive signal. Furthermore, samples will be considered Heterozygous of wild and mutant alleles whenever they display a VIC and FAM positive signal.

2.1 Statistical analysis

Data were analyzed with SPSS v25.0. Group comparisons of allele and genotype frequencies were performed using linear logistic regression. The relationship between COVID-19 severity and the IFNL4 gene (rs12979860) was determined with multiple logistic regression (linear generalized model). Statistical significance was set at $p \leq 0.05$.

3. Result

Patients with mild-moderate infection showed a lower mean age (45.58 ± 15.96 years) than those with severe infection (57.73 ± 10.85 years) with highly significant differences ($p < 0.001$), females were more frequent in both group (54.67% vs. 52.00%), the difference was not significant ($p < 0.56$) Table 1.

The wild homozygous genotype (CC) was more frequent among mild/moderate group than severe group (73.33% vs. 52.33%) with a significant difference. In contrast, the heterozygous genotype (TC) and mutant homozygous genotype (TT) were more common among Severe group (29.33% and 18.33%, respectively) than mild/moderate group (18.6% and 8.00%, respectively) with significant differences ($OR = 0.45$, $95\%CI = 0.19-1.06$, and $OR = 0.30$, $95\%CI = 0.09-0.94$, $p = 0.021$, respectively).

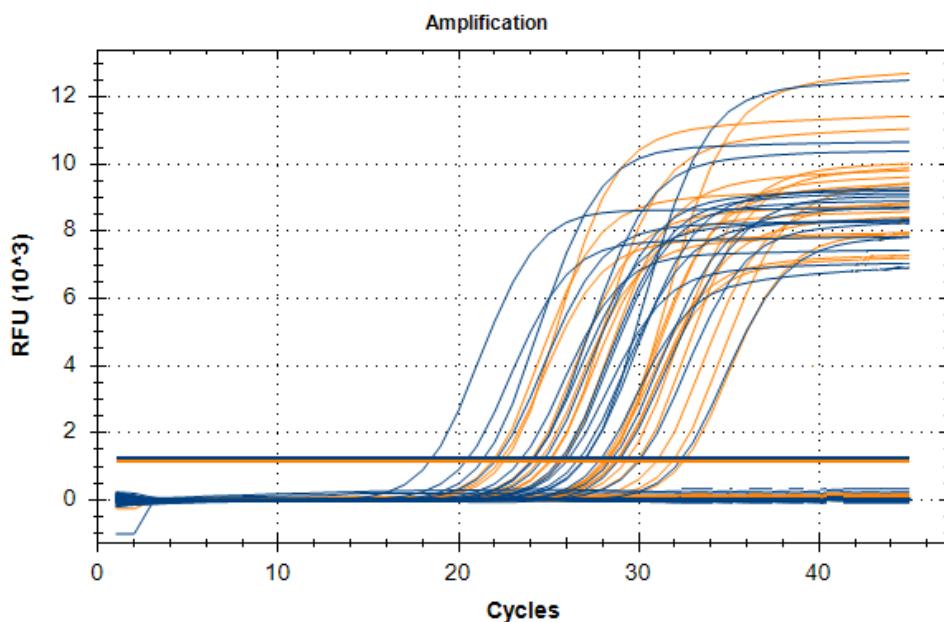
Analysis of allele distribution (Figure1) revealed a higher frequency of T allele among severe group than mild/moderate group (33.33% versus 17.33%) with a highly significant difference ($OR = 0.27$, $95\%CI = 0.10-0.64$, $p = 0.002$) Table 2.

Table 1. Demographic characteristics of the study population.

Factor	Mild-moderate	Severe	P value (sig≤0.05)
Age, years			
Mean ±SD	45.58±15.96	57.73±10.85	<0.001
Gender			
Male	35(45.33%)	48(48.00%)	0.56
Female	45(54.67%)	52(52.00%)	

Table 2. Variations of IFNL4 rs12979860 in the COVID-19 patients.

rs12979860	Mild-moderate	Severe	P value (sig≤0.05)	OR (95%CI)
Genotypes				
CC	55(73.33%)	39(52.00%)		1.0
CT	14(18.66%)	22(29.33%)	0.021*	0.45(0.19-1.06)
TT	6(8.00%)	14(18.66%)		0.30(0.09-0.94)
Alleles				
C	124(82.66%)	100(66.66%)		1.0
T	26(17.33%)	50(33.33%)	0.002*	0.27(0.10-0.64)

**Figure 1.** Genotyping of Alleles using TaqMan Real time PCR the blue line represent T Allele and orange line represent C Allele ([Prepared by Authors, 2025](#)).

4. Discussion

According to our results, TT genotype is significantly expressed in COVID-19 sever patients versus mild/moderate patients. Also, T allele is higher risk of COVID-19 severity. Many associations have been found between interferon lambda 4 rs12979860 polymorphism and various clinical outcomes such as viral clearance of HCV and other RNA viruses (26-28). The association between the polymorphism in the IFN-lambda gene (rs12979860) and COVID-19 has been reported in many papers (24, 25, 29-36). In this sense, a study done in Spain by Saponi-Cortes et al (24), the T allele of rs12979860 was more prevalent in COVID-19 patients than in healthy controls, indicating its potential role as a risk factor for infection (24). Furthermore, another study that included 750 SARS-CoV-2 positive patients (375 survivors and 375 nonsurvivors) disclosed that the results of that study proved that the severity of COVID-19 infection was associated with unfavorable genotypes of IFNL4 rs1297860 (TT) (25). In another study done by Zahid et al (37) included 117 subjects found that T allele more frequent in sever patients compared to mild patients with no statistical difference (37). Agwa et al (38) remarked that rs12979860 can predispose to COVID-19 and they mentioned that the highest frequency of T allele of rs12979860 was more frequent in COVID-19 patients with regard to the general population (36.16% vs. 26.40%) so they conclude that the rs12979860 polymorphism of IFNL4 can predispose to COVID-19 (38). According to the results in this study as well as the other results it's revealed that the C allele of rs12252 and T allele of rs12979860 are related to the poor outcome of the disease, which may be due to producing nonfunctional protein. However, it's essential to note that the association between IFNL4 rs12979860 T polymorphism and COVID-19 outcomes is still under study, and results may vary between different populations and studies.

5. Conclusion

We conclude that TT variation as well as T allele are associated with the severity of COVID-19 infection and the severity of disease was positively associated with advancing age among patients' groups. IFNL4, a type III interferon, is critical for antiviral defense in epithelial tissues. The rs12979860 polymorphism influences viral clearance and disease outcomes, with the T allele associated with increased risk of severe COVID-19, while the C allele may offer protective effects.

6. Declarations

References

1. Zhu N, Zhang D, Wang W, Li X, Yang B, Song J, et al. A Novel Coronavirus from Patients with Pneumonia in China, 2019. *N Engl J Med*. 2020;382(8):727-33. [\[PMID\]](#) [\[PMCID\]](#) [\[DOI:10.1056/NEJMoa2001017\]](#)

6.1 Acknowledgments

None.

6.2 Ethical Considerations

This study was carried out under the ethical approval of the institutional board review (IRB) in the College of Medicine Al-Nahrain University, under the number 20211063.

6.3 Authors' Contributions

Conceptualization and Design: Sara F. Ahmed and Muhammed A. H. Aldabagh. Data Acquisition: Sara F. Ahmed and Aryan F. Khorshid. Analysis and Interpretation: Sara F. Ahmed. Methodology: Sara F. Ahmed and Muhammed A. H. Aldabagh. Writing: Sara F. Ahmed. Supervision: Sara F. Ahmed and Muhammed A. H. Aldabagh. Resources: Aryan F. Khorshid. Data Curation: Sara F. Ahmed. All authors reviewed, edited, and approved the final version of the manuscript.

6.4 Conflict of Interest

The authors declare no conflict of interest.

6.5 Fund or Financial Support

This research received no specific grant from any funding agency in the public, commercial, or not-for-profit sectors.

6.6 Using Artificial Intelligence Tools (AI Tools)

The authors were not utilized AI Tools.

7. Publisher's Note

This article is part of the Special Issue arising from the Second International Conference for Pharmaceutical Sciences (SICPS 2025), College of Pharmacy, University of Misan, Iraq (29 Nov–1 Dec 2025, see <https://uomisan.edu.iq/pharmacy/conference/>). All manuscripts in this issue were peer-reviewed and accepted for publication in *Journal of Advances in Medical and Biomedical Research (J Adv Med Biomed Res)*.

2. Ali AM, Tofiq AM, Rostam HM, Ali KM, Tawfeeq HM. Disease severity and efficacy of homologous vaccination among patients infected with SARS-CoV-2 Delta or Omicron VOCs, compared to unvaccinated using main biomarkers. *J Med Virol.* 2022;94(12):5867-76. [\[DOI:10.1002/jmv.28098\]](https://doi.org/10.1002/jmv.28098) [\[PMID\]](https://pubmed.ncbi.nlm.nih.gov/35900000/) [\[PMCID\]](https://pubmed.ncbi.nlm.nih.gov/35900000/)
3. Vabret N, Britton GJ, Gruber C, Hegde S, Kim J, Kuksin M, et al. Immunology of COVID-19: Current State of the Science. *Immunity.* 2020; 52(6):910-41. [\[PMCID\]](https://pubmed.ncbi.nlm.nih.gov/32500000/) [\[DOI:10.1016/j.immuni.2020.05.002\]](https://doi.org/10.1016/j.immuni.2020.05.002) [\[PMID\]](https://pubmed.ncbi.nlm.nih.gov/32500000/)
4. Webb Hooper M, Napoles AM, Perez-Stable EJ. COVID-19 and Racial/Ethnic Disparities. *JAMA.* 2020;323(24):2466-7. [\[PMCID\]](https://pubmed.ncbi.nlm.nih.gov/32500000/) [\[DOI:10.1001/jama.2020.8598\]](https://doi.org/10.1001/jama.2020.8598) [\[PMID\]](https://pubmed.ncbi.nlm.nih.gov/32500000/)
5. Mehta P, McAuley DF, Brown M, Sanchez E, Tattersall RS, Manson JJ. COVID-19: consider cytokine storm syndromes and immunosuppression. *Lancet.* 2020; 395(10229):1033-4. [\[PMID\]](https://pubmed.ncbi.nlm.nih.gov/32500000/) [\[PMCID\]](https://pubmed.ncbi.nlm.nih.gov/32500000/) [\[DOI:10.1016/S0140-6736\(20\)30628-0\]](https://doi.org/10.1016/S0140-6736(20)30628-0)
6. Xu Z, Shi L, Wang Y, Zhang J, Huang L, Zhang C, et al. Pathological findings of COVID-19 associated with acute respiratory distress syndrome. *Lancet Respir Med.* 2020; 8(4):420-2. [\[PMID\]](https://pubmed.ncbi.nlm.nih.gov/32500000/) [\[PMCID\]](https://pubmed.ncbi.nlm.nih.gov/32500000/) [\[DOI:10.1016/S2213-2600\(20\)30076-X\]](https://doi.org/10.1016/S2213-2600(20)30076-X)
7. Rockx B, Kuiken T, Herfst S, Bestebroer T, Lamers MM, Oude Munnink BB, et al. Comparative pathogenesis of COVID-19, MERS, and SARS in a nonhuman primate model. *Science.* 2020;368(6494):1012-5. [\[DOI:10.1126/science.abb7314\]](https://pubmed.ncbi.nlm.nih.gov/32500000/) [\[PMID\]](https://pubmed.ncbi.nlm.nih.gov/32500000/) [\[PMCID\]](https://pubmed.ncbi.nlm.nih.gov/32500000/)
8. Moore JB, June CH. Cytokine release syndrome in severe COVID-19. *Science.* 2020; 368(6490):473-4. [\[DOI:10.1126/science.abb8925\]](https://pubmed.ncbi.nlm.nih.gov/32500000/) [\[PMID\]](https://pubmed.ncbi.nlm.nih.gov/32500000/)
9. 9Lei J, Li J, Li X, Qi X. CT Imaging of the 2019 Novel Coronavirus (2019-nCoV) Pneumonia. *Radiology.* 2020;295(1):18. [\[DOI:10.1148/radiol.2020200236\]](https://pubmed.ncbi.nlm.nih.gov/32500000/) [\[PMID\]](https://pubmed.ncbi.nlm.nih.gov/32500000/) [\[PMCID\]](https://pubmed.ncbi.nlm.nih.gov/32500000/)
10. Ovsyannikova IG, Haralambieva IH, Crooke SN, Poland GA, Kennedy RB. The role of host genetics in the immune response to SARS-CoV-2 and COVID-19 susceptibility and severity. *Immunol Rev.* 2020;296(1):205-19. [\[DOI:10.1111/imr.12897\]](https://pubmed.ncbi.nlm.nih.gov/32500000/) [\[PMID\]](https://pubmed.ncbi.nlm.nih.gov/32500000/) [\[PMCID\]](https://pubmed.ncbi.nlm.nih.gov/32500000/)
11. Ali HN, Ali KM, Rostam HM, Ali AM, Tawfeeq HM, Fatah MH, et al. Clinical laboratory parameters and comorbidities associated with severity of coronavirus disease 2019 (COVID-19) in Kurdistan Region of Iraq. *Pract Lab Med.* 2022;31:e00294. [\[PMID\]](https://pubmed.ncbi.nlm.nih.gov/35900000/) [\[DOI:10.1016/j.plabm.2022.e00294\]](https://doi.org/10.1016/j.plabm.2022.e00294) [\[PMCID\]](https://pubmed.ncbi.nlm.nih.gov/35900000/)
12. Velavan TP, Pallerla SR, Ruter J, Augustin Y, Kremsner PG, Krishna S, et al. Host genetic factors determining COVID-19 susceptibility and severity. *EBioMedicine.* 2021;72:103629. [\[DOI:10.1016/j.ebiom.2021.103629\]](https://doi.org/10.1016/j.ebiom.2021.103629) [\[PMID\]](https://pubmed.ncbi.nlm.nih.gov/35900000/) [\[PMCID\]](https://pubmed.ncbi.nlm.nih.gov/35900000/)
13. Yu K, Wang J, Li H, Wang W. IFITM3 rs12252 polymorphism and coronavirus disease 2019 severity: A meta-analysis. *Exp Ther Med.* 2023;25(4):158. [\[PMCID\]](https://pubmed.ncbi.nlm.nih.gov/35900000/) [\[DOI:10.3892/etm.2023.11857\]](https://doi.org/10.3892/etm.2023.11857) [\[PMID\]](https://pubmed.ncbi.nlm.nih.gov/35900000/)
14. Gupta K, Kaur G, Pathak T, Banerjee I. Systematic review and meta-analysis of human genetic variants contributing to COVID-19 susceptibility and severity. *Gene.* 2022;844:146790. [\[DOI:10.1016/j.gene.2022.146790\]](https://doi.org/10.1016/j.gene.2022.146790) [\[PMID\]](https://pubmed.ncbi.nlm.nih.gov/35900000/) [\[PMCID\]](https://pubmed.ncbi.nlm.nih.gov/35900000/)
15. Pecoraro V, Cuccorese M, Trenti T. Genetic polymorphisms of ACE1, ACE2, IFITM3, TMPRSS2 and TNFalpha genes associated with susceptibility and severity of SARS-CoV-2 infection: a systematic review and meta-analysis. *Clin Exp Med.* 2023;23(7):3251-64. [\[DOI:10.1007/s10238-023-01038-9\]](https://doi.org/10.1007/s10238-023-01038-9) [\[PMID\]](https://pubmed.ncbi.nlm.nih.gov/35900000/) [\[PMCID\]](https://pubmed.ncbi.nlm.nih.gov/35900000/)
16. Sommereyns C, Paul S, Staeheli P, Michiels T. IFN-lambda (IFN-lambda) is expressed in a tissue-dependent fashion and primarily acts on epithelial cells in vivo. *PLoS Pathog.* 2008; 4(3):e1000017. [\[PMID\]](https://pubmed.ncbi.nlm.nih.gov/35900000/) [\[PMCID\]](https://pubmed.ncbi.nlm.nih.gov/35900000/) [\[DOI:10.1371/journal.ppat.1000017\]](https://doi.org/10.1371/journal.ppat.1000017)
17. Mordstein M, Neugebauer E, Ditt V, Jessen B, Rieger T, Falcone V, et al. Lambda interferon renders epithelial cells of the respiratory and gastrointestinal tracts resistant to viral infections. *J Virol.* 2010;84(11):5670-7. [\[DOI:10.1128/JVI.00272-10\]](https://doi.org/10.1128/JVI.00272-10) [\[PMID\]](https://pubmed.ncbi.nlm.nih.gov/35900000/) [\[PMCID\]](https://pubmed.ncbi.nlm.nih.gov/35900000/)
18. Baldridge MT, Lee S, Brown JJ, McAllister N, Urbanek K, Dermody TS, et al. Expression of Ifnlr1 on Intestinal Epithelial Cells Is Critical to the Antiviral Effects of Interferon Lambda against Norovirus and Reovirus. *J Virol.* 2017; 91(7):e02079. [\[DOI:10.1128/JVI.02079-16\]](https://doi.org/10.1128/JVI.02079-16) [\[PMID\]](https://pubmed.ncbi.nlm.nih.gov/35900000/) [\[PMCID\]](https://pubmed.ncbi.nlm.nih.gov/35900000/)
19. Kotenko SV, Gallagher G, Baurin VV, Lewis-Antes A, Shen M, Shah NK, et al. IFN-lambdas mediate antiviral protection through a distinct class II cytokine receptor complex. *Nat Immunol.* 2003;4(1):69-77. [\[DOI:10.1038/ni875\]](https://doi.org/10.1038/ni875) [\[PMID\]](https://pubmed.ncbi.nlm.nih.gov/35900000/)
20. Sharafi H, Alavian SM, Behnava B, Pouryasin A, Keshvari M. The Impact of IFNL4 rs12979860 Polymorphism on Spontaneous

Clearance of Hepatitis C; A Case-Control Study. *Hepat Mon.* 2014;14(10):e22649. [\[DOI:10.5812/hepatmon.22649\]](https://doi.org/10.5812/hepatmon.22649)

21. Tanaka Y, Nishida N, Sugiyama M, Kurosaki M, Matsuura K, Sakamoto N, et al. Genome-wide association of IL28B with response to pegylated interferon-alpha and ribavirin therapy for chronic hepatitis C. *Nat Genet.* 2009;41(10):1105-9. [\[DOI:10.1038/ng.449\]](https://doi.org/10.1038/ng.449) [\[PMID\]](#)
22. Lapinski TW, Pogorzelska J, Kowalcuk O, Niklinski J, Flisiak R. SNP RS12979860 related spontaneous clearance of hepatitis c virus infection in HCV/HIV-1 coinfecting patients. *Przegl Epidemiol.* 2013;67(3):407-9, 517-9.
23. Farzanegan Gharabolagh A, Bamdad T, Hedayati M, Dehghan Manshadi SA. The Synergistic Effect of Fluvastatin and IFN-lambda on Peripheral Blood Mononuclear Cells of Chronic Hepatitis C Virus (HCV) Patients with IL-28B rs12979860 CC Genotype. *Iran J Allergy Asthma Immunol.* 2019;18(5):533-42. [\[DOI:10.18502/ijaa.v18i5.1923\]](https://doi.org/10.18502/ijaa.v18i5.1923) [\[PMID\]](#)
24. Saponi-Cortes JMR, Rivas MD, Calle-Alonso F, Sanchez JF, Costo A, Martin C, et al. IFNL4 genetic variant can predispose to COVID-19. *Sci Rep.* 2021;11(1):21185. [\[PMCID\]](#) [\[DOI:10.1038/s41598-021-00747-z\]](https://doi.org/10.1038/s41598-021-00747-z) [\[PMID\]](#)
25. Rahimi P, Tarharoudi R, Rahimpour A, Mosayebi Amroabadi J, Ahmadi I, Anvari E, et al. The association between interferon lambda 3 and 4 gene single-nucleotide polymorphisms and the recovery of COVID-19 patients. *Virol J.* 2021;18(1):221. [\[PMID\]](#) [\[PMCID\]](#) [\[DOI:10.1186/s12985-021-01692-z\]](https://doi.org/10.1186/s12985-021-01692-z)
26. Prokunina-Olsson L, Muchmore B, Tang W, Pfeiffer RM, Park H, Dickensheets H, et al. A variant upstream of IFNL3 (IL28B) creating a new interferon gene IFNL4 is associated with impaired clearance of hepatitis C virus. *Nat Genet.* 2013;45(2):164-71. [\[PMID\]](#) [\[PMCID\]](#) [\[DOI:10.1038/ng.2521\]](https://doi.org/10.1038/ng.2521) [\[PMID\]](#) [\[PMCID\]](#)
27. Thomas DL, Thio CL, Martin MP, Qi Y, Ge D, O'Huigin C, et al. Genetic variation in IL28B and spontaneous clearance of hepatitis C virus. *Nature.* 2009;461(7265):798-801. [\[PMID\]](#) [\[PMCID\]](#) [\[DOI:10.1038/nature08463\]](https://doi.org/10.1038/nature08463) [\[PMID\]](#) [\[PMCID\]](#)
28. Rugwizangoga B, Andersson ME, Kabayiza JC, Nilsson MS, Armannsdottir B, Aurelius J, et al. IFNL4 Genotypes Predict Clearance of RNA Viruses in Rwandan Children With Upper Respiratory Tract Infections. *Front Cell Infect Microbiol.* 2019;4(9):340. [\[PMCID\]](#) [\[DOI:10.3389/fcimb.2019.00340\]](https://doi.org/10.3389/fcimb.2019.00340) [\[PMID\]](#)
29. Amadio E, Pipitone RM, Grimaudo S, Immordino P, Maida CM, Prestileo T, et al. SARS-CoV-2 Viral Load, IFNlambda Polymorphisms and the Course of COVID-19: An Observational Study. *J Clin Med.* 2020;9(10):3315. [\[DOI:10.3390/jcm9103315\]](https://doi.org/10.3390/jcm9103315) [\[PMID\]](#) [\[PMCID\]](#)
30. Zhang Y, Qin L, Zhao Y, Zhang P, Xu B, Li K, et al. Interferon-Induced Transmembrane Protein 3 Genetic Variant rs12252-C Associated With Disease Severity in Coronavirus Disease 2019. *J Infect Dis.* 2020;222(1):34-7. [\[DOI:10.1093/infdis/jiaa224\]](https://doi.org/10.1093/infdis/jiaa224) [\[PMID\]](#) [\[PMCID\]](#)
31. Alghamdi J, Alaamery M, Barhoumi T, Rashid M, Alajmi H, Aljasser N, et al. Interferon-induced transmembrane protein-3 genetic variant rs12252 is associated with COVID-19 mortality. *Genomics.* 2021;113(4):1733-41. [\[DOI:10.1016/j.ygeno.2021.04.002\]](https://doi.org/10.1016/j.ygeno.2021.04.002) [\[PMID\]](#) [\[PMCID\]](#)
32. Cuesta-Llavona E, Albaiceta GM, Garcia-Clemente M, Duarte-Herrera ID, Amado-Rodriguez L, Hermida-Valverde T, et al. Association between the interferon-induced transmembrane protein 3 gene (IFITM3) rs34481144 / rs12252 haplotypes and COVID-19. *Curr Res Virol Sci.* 2021;2:100016. [\[DOI:10.1016/j.crviro.2021.100016\]](https://doi.org/10.1016/j.crviro.2021.100016) [\[PMID\]](#) [\[PMCID\]](#)
33. Gomez J, Albaiceta GM, Cuesta-Llavona E, Garcia-Clemente M, Lopez-Larrea C, Amado-Rodriguez L, et al. The Interferon-induced transmembrane protein 3 gene (IFITM3) rs12252 C variant is associated with COVID-19. *Cytokine.* 2021;137:155354. [\[DOI:10.1016/j.cyto.2020.155354\]](https://doi.org/10.1016/j.cyto.2020.155354) [\[PMID\]](#)
34. Schonfelder K, Breuckmann K, Elsner C, Dittmer U, Fistera D, Herbstreit F, et al. The influence of IFITM3 polymorphisms on susceptibility to SARS-CoV-2 infection and severity of COVID-19. *Cytokine.* 2021;142:155492. [\[DOI:10.1016/j.cyto.2021.155492\]](https://doi.org/10.1016/j.cyto.2021.155492) [\[PMID\]](#) [\[PMCID\]](#)
35. Mulla S, Molla MMA, Ahmed SMA, Akhtaruzzaman AKM, Saleh AA, Anwar S. Association of interferon gamma inducible protein-10, monocyte chemoattractant protein-1, macrophage inflammatory protein-1 alpha, interleukin-6, and rs12252 single nucleotide polymorphism of interferon-induced transmembrane protein-3 gene with the severity of COVID-19 infection. *Egypt J Intern Med.* 2022;34(1):53. [\[PMID\]](#) [\[PMCID\]](#) [\[DOI:10.1186/s43162-022-00141-9\]](https://doi.org/10.1186/s43162-022-00141-9)
36. Ahmadi I, Afifipour A, Sakhaee F, Zamani MS, Mirzaei Gheinari F, Anvari E, et al. Impact

of interferon-induced transmembrane protein 3 gene rs12252 polymorphism on COVID-19 mortality. *Cytokine*. 2022;157:155957. [PMID] [DOI:10.1016/j.cyto.2022.155957] [PMCID]

37. Zahid W, Farooqui N, Zahid N, Ahmed K, Anwar MF, Rizwan-Ul-Hasan S, et al. Association of Interferon Lambda 3 and 4 Gene SNPs and Their Expression with COVID-19 Disease Severity: A Cross-Sectional Study. *Infect Drug Resist*. 2023;16:6619-28. [PMID] [DOI:10.2147/IDR.S422095] [PMCID]

38. Agwa SHA, Kamel MM, Elghazaly H, Abd Elsamee AM, Hafez H, Girgis SA, et al. Association between Interferon-Lambda-3 rs12979860, TLL1 rs17047200 and DDR1 rs4618569 Variant Polymorphisms with the Course and Outcome of SARS-CoV-2 Patients. *Genes (Basel)*. 2021;12(6):830. [PMCID] [DOI:10.3390/genes12060830] [PMID]

How to Cite This Article:

Ahmed S F, Aldabagh M A H, Khorshid A F, Hawezy D J. The Association of Interferon Lambda 4 (IFNL4) rs12979860 Polymorphism with COVID-19 Outcome. *J Adv Med Biomed Res*. 2025;33(162):81-7.

Download citation:

[BibTeX](#) | [RIS](#) | [EndNote](#) | [Medlars](#) | [ProCite](#) | [Reference Manager](#) | [RefWorks](#)

Send citation to:

 [Mendeley](#)  [Zotero](#)  [RefWorks](#)